



User Guide



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What is AbGenesis?

AbGenesis is an analysis platform that allows scientists to manage, analyze and share results from antibody sequencing experiments. AbGenesis provides scientists access to unique Distributed Bio algorithms that will accelerate your antibody research. It is a lightweight highly customizable application that is tailored to your scientists' specific workflows and analysis needs.

abgenesis.distributedbio.org shows a snapshot of how the system could be deployed. For a full demo or discussion of how you could use AbGenesis contact info@distributedbio.com

Browser Information

To use AbGenesis you will need a web browser. We recommend either Firefox or Chrome. Both are free and are simple to install on most operating systems. Get Firefox at http://www.mozilla.org and Chrome at http://www.google.com/chrome

The latest version of Internet Explorer will work with the system, but some data rich pages will load more slowly.

Cookies must be enabled. To enable cookies follow the instructions here:

Firefox http://support.mozilla.org/en-US/kb/enable-and-disable-cookies-website-

preferences

Chrome http://support.google.com/accounts/bin/answer.py?hl=en&answer=61416

To use the Jalview alignment viewer that is embedded in the system Java must be enabled.

Important Concepts in the System

User an individual user of the system, can upload sequences and browse

results

Group a group of users who can all see the same Projects and Experiments,

typically all users from the same company are in the same group

Project a container for multiple experiments that relate to the same goal

Experiment a set of sequences to be analyzed



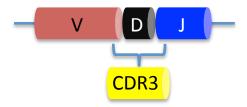
The Relationship Between Sequences, Clones and Lineages

You give us DNA sequences:

>lgG1

TGACGTGGGAAGACCGATGGGCCCTTGGTGGAGGCT......

where possible for each sequence we identify a V,D & J gene, CDRs1, 2 & 3



IGHV4-4, IGHD3-22, IGHJ4, GSITNYCWS, GRIYPSGYTNYS, CARVLYDSCGYYHFDYW

CLONE - we identify a clone as the V & J genes and the CDR3



IGHV4-4 IGHJ4 CARVLYDSCGYYHFDYW

Representative Sequence – if multiple sequences can have the same V/J CDR3 combination we assign 1 sequence to represent this clone. The Representative Sequence will be used in Developability analysis and will be shown in all reports.

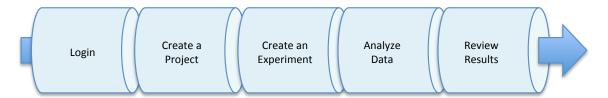
Lineages are groups of clones that have the same V & J, and CDR3s that differ by a max of 2 amino acids. A lineage is identified by the first clone in the group, e.g. our clone from above can be found in the lineage below

IGHV4-4_IGHJ4_CARVLYDSRGYYHFDYW IGHV4-4 IGHJ4 CARVLYDSCGYYHFDYW



General Workflow

The overall workflow is as follows:

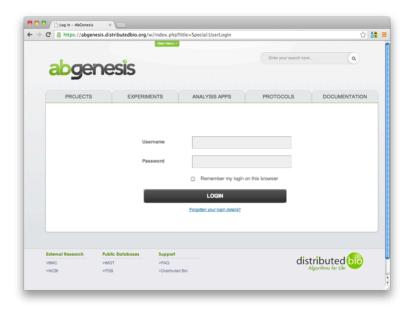


Once you have created a Project you can of course create as many Experiments as you want in it. You should remember that it is best to create Projects where the Experiments or pertain to the same goal. That way some of the analysis AbGenesis presents will be more useful.

Step 1 – Login in to the system

Once you have a working web browser type http://abgenesis.distributedbio.org into the URL box.

You will briefly see the landing page and then this page will load

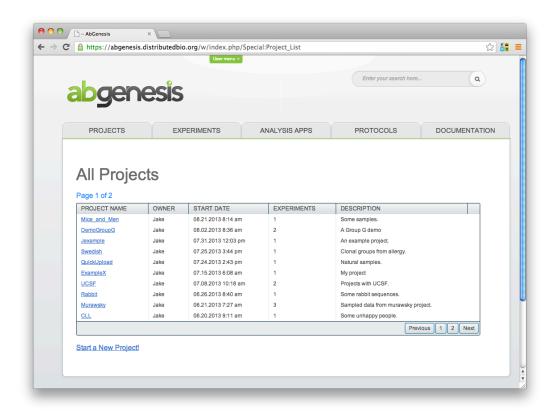


Fill in the username and password you have been provided by Distributed Bio and press the Log In button. To get an account on the system contact your Distributed Bio representative.



support@distributedbio.com

When you have logged in this the Projects Home Page will load (below). If no one in your group has created a Project before you will not see any Projects listed.



Navigation Bar

At the top of all AbGenesis pages is the Navigation Bar. It contains useful links that will help you rapidly navigate the system.



PROJECTS A list of the Projects your Group have created

EXPERIMENTS A list of Experiments from all Projects your Group has created

ANALYSIS APPS A page where you can launch individual analysis runs

PROTOCOLS A page where you can store information on Protocols your Group

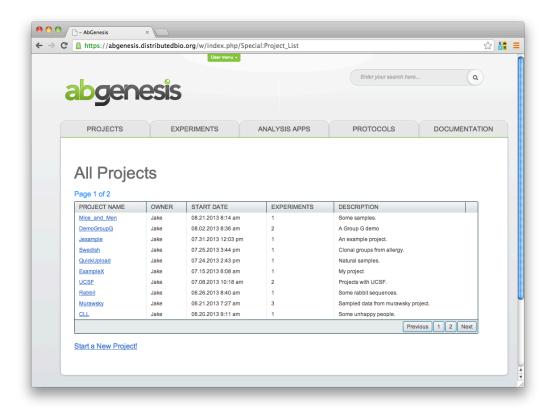
uses

DOCUMENTATION Useful documentation



Step 2 – Create a Project

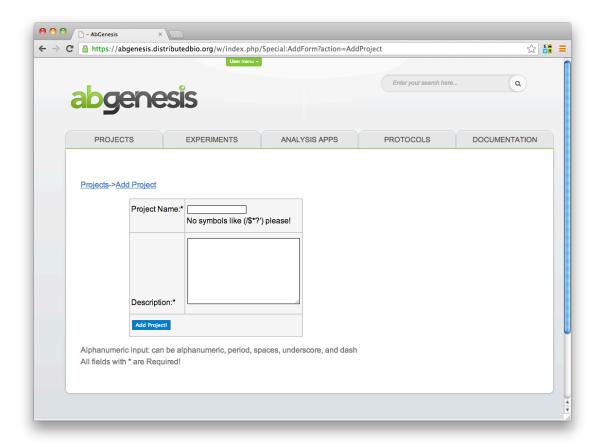
The Projects Home Page shown below shows the Project Name, the Owner and Start Date, Number of Experiments in the Project and the Description. At any time if you press the Projects navigation bar link you will come back here.



To create a new Project press the "Start a New Project" link at the bottom of the table.



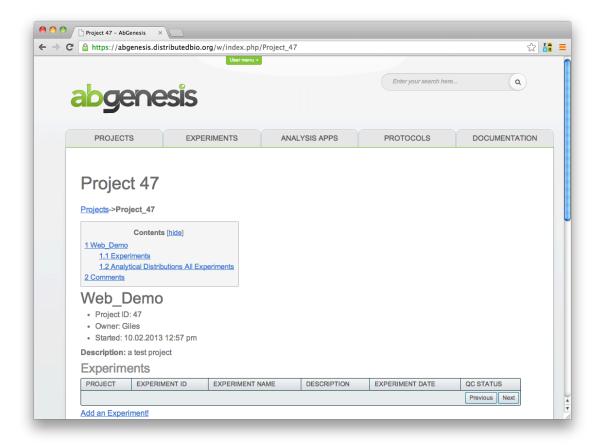
This page will load:



Fill in the form and press the "Add Project" button.



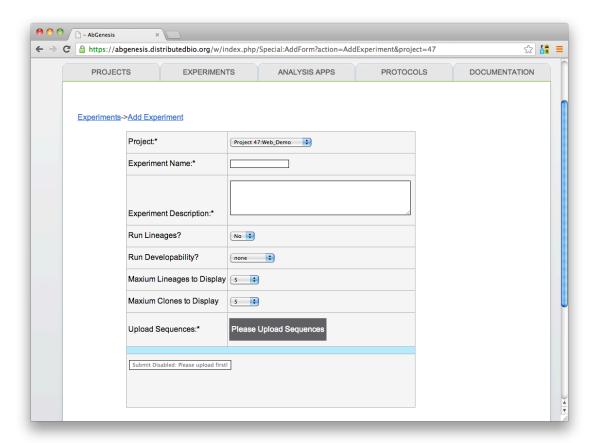
This page will now load showing a summary of what you just entered. The Project number (11 below) is automatically created by the system and is the unique identifier for this Project.





Step 3 - Create an Experiment

Click the "Add an Experiment" button at the bottom of the Experiments Table. This page will load:



Enter an Experiment Name and Description.

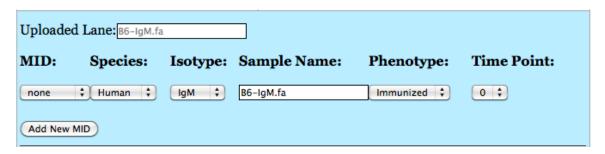
Options

Run Lineages	Calculate clone trees based VDJ segments	
Run Developability	Calculate bio-physical characteristics that make	
	preferred therapeutic antibodies	
Maximum Lineages to Display	If you are running 1000s of sequences you can't display	
	them all in the report – limit the display to this	
Maximum Clones to Display	If you are running 1000s of sequences you can't display	
	them all in the report — limit the display to this	

Next you need to add some sequence files. You can either drag and drop these onto the "Please Upload Sequences" button, or press the button to activate a file chooser. See Appendix A for acceptable file formats to upload. The Submit button will not be activated until you have chosen some files.



When the files have uploaded a box like this will appear for every file you uploaded:



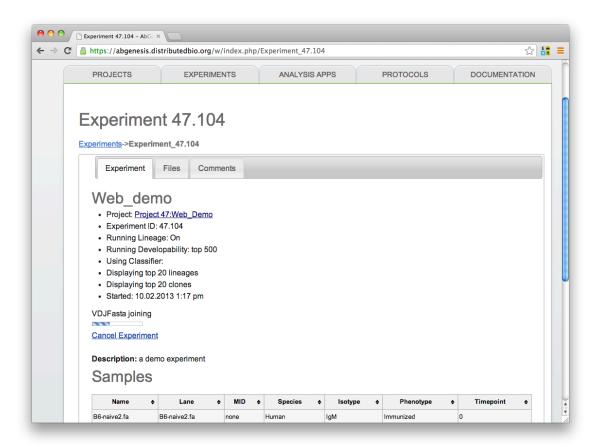
If you uploaded multiple files there will be multiple boxes stacked on top of each other. These boxes allow you to MID and Phenotype details for the sequences contained in that file

Once you have finished adding the extra detail press the "Lets Go" button at the bottom of the screen.

The system will now start analyzing your experiments. This may take a few minutes depending on the number of sequences you uploaded.



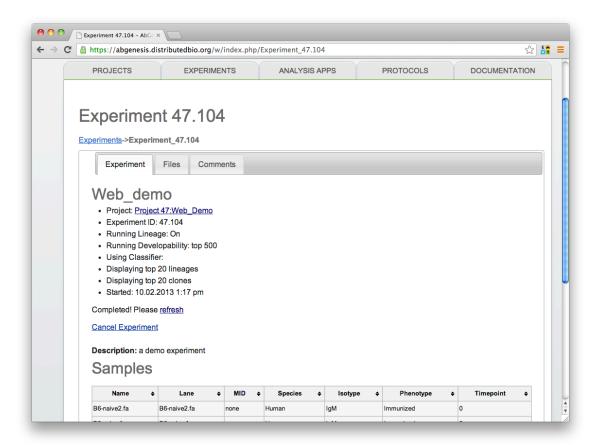
You should see a screen that looks like this:



The Progress Bar seen here in the middle of the screen will update you on the progress of your run. You can leave this page without affecting the completion of your analysis. You can return to the results by clicking through to this Experiment from the Project Home page.



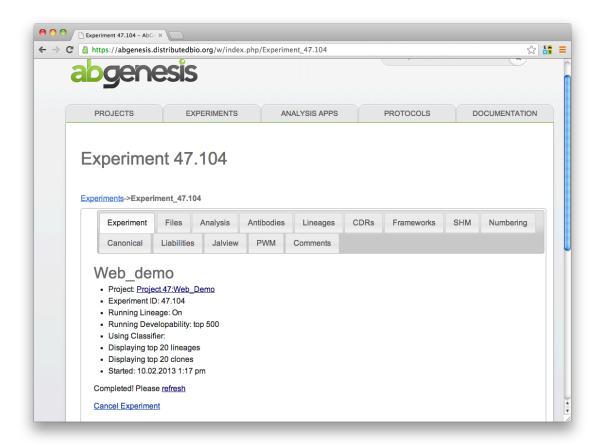
When your analysis has complete you will see a screen that looks like this:



The Progress Bar will have changed into a "refresh" link. Click on this to view to your results.



The screen should change to something that looks like this:



You will now be able to review your results by clicking through the tabs displayed. See Step 4 for details of what all the data is on each tab.

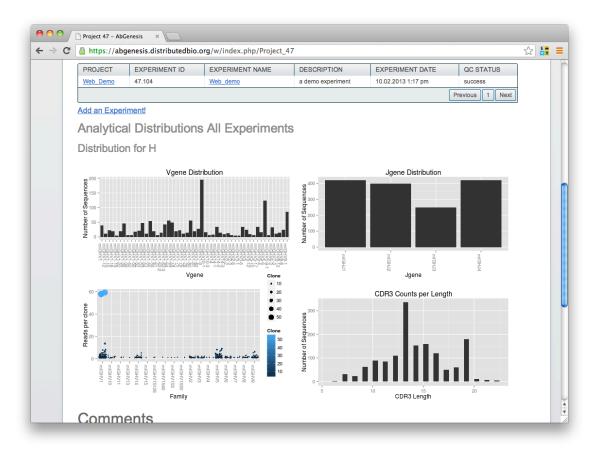


Step 4 - Review Data

To see the results of the Experiment either click the refresh link on Experiments page that was generated or click through to the Experiment from the Project Summary page.

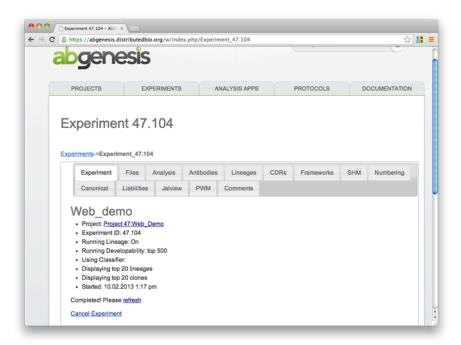
Project Summary Page

As shown below will give some summary statistics collated from all Experiments in a Project.



Click on the Experiment Name for the Experiment you are interested in to load the Experiment page as shown below:





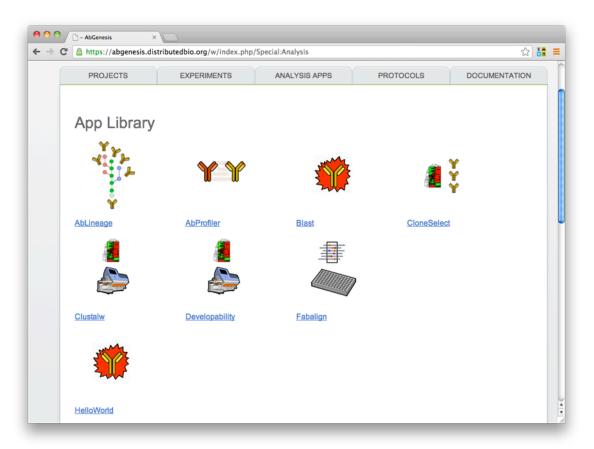
The Experiment page contains the results of the analysis pipeline for the sequences in that file as described below:

Tab	Description
Experiment	Summary of the experiment options chosen
Files	A list of the files uploaded and files generated by the analysis
Analysis	Summary of results across the sequences including: read length;
	depth; orientation; on-target QC; cross-sample clones & SHM Profiles
	grouped by input file
Antibodies	Analysis of each sequence for V&J gene distribution
Lineages	Sequences with the same V,J and same composition of CDR3 (allowed
	to be up to 2 residues different) are the same Lineage. Click on the
	Lineage to see the sequences that contain it.
CDRs	CDR sequences for each sequence
Frameworks	List of matched frameworks
SHM	Somatic Hypermutations found in the sequences
Numbering	Kabat numerbing for each sequence — click on each sequence to see
	the numbering for that sequence
Canonical	The MLT canonical classification of the CDRs. CDR-3 labled as kinked
	or un-kinked depending on whether the salt-bridge is intact.
Liabilities	Biochemical liabilities for the sequence: acid hydrolysis sites; splice
	sites; cryptic splice sites; N-linked glycosylation sites; methionines.
Jalview	Java applet aligning all the sequences in an interactive window
PWM	Positional Weight Matrices for CDRs in VH, VK & VL
Comments	A tab for Users to add comments on the Experiment



Analysis Apps

There may be several analysis tools deployed on your system that you can use to run an analysis that you do not want to add to a Project or Experiment. Click on the ANALYSIS APPS tab from the Navigation Bar to see them.



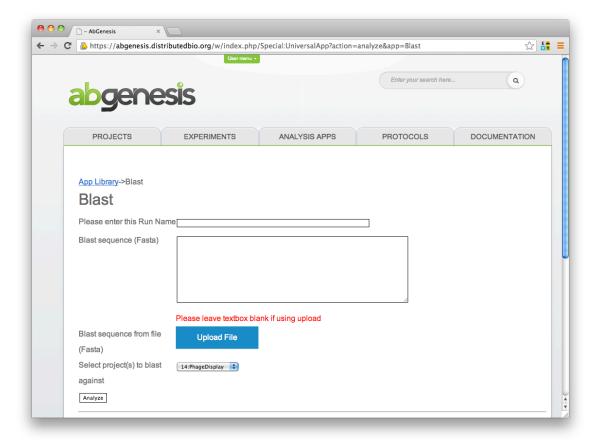
The 4 analysis app icons are shown, plus some information on latest runs and current server activity.

App	Description
Blast	Blast a single sequence against sequences uploaded to the system
Developability	Analyses sequences for various developability features and abnormalities.
Fabalign	Generates a Hidden Markov Model from input fasta amino acid sequences with cdr kabat-restacking.
Humanize	Find the best humanization scaffolds



Blast

This allows you to perform a single Blast search against any sequence from a Project. Give the run a name and paste in a Fasta (see Appendix A) sequence into the Query box. You can choose an individual Project from the drop down menu, or select All to Blast all sequences.

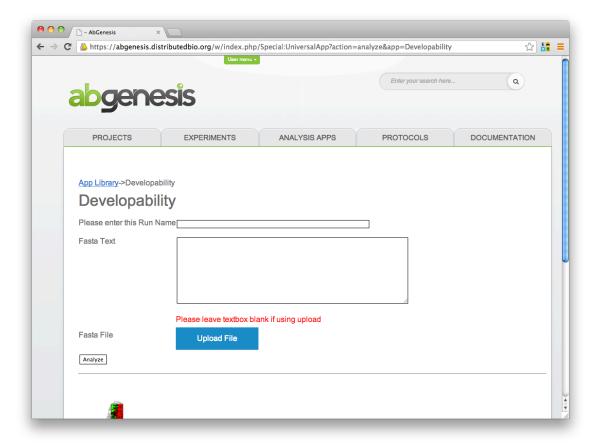


When you have picked a database click Analyse.



Developability

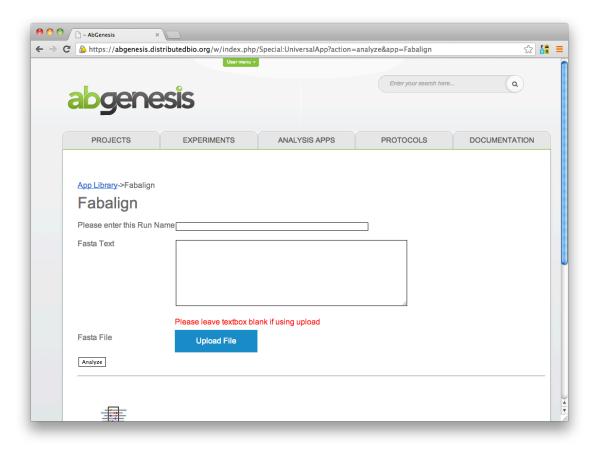
This will generate most of the features generated for an Experiment for a group of sequences you do not want to add to a Project or Experiment. Give a name for you run and paste in 1 or more fasta (see Appendix A) nucleotide sequences. Click Analyse.





Fabalign

Given the amino acid sequence of an antibody heavy or light chain will generate a multiple sequence alignment with CDR Kabat re-stacking. Give your urn a name and paste in a fasta query sequence and click Analyze.





Appendix A - Supported Sequence File Formats

- Sanger, MiSeq, HiSeq, Ion torrent, 454
- Fasta, fastq, Sanger Zip, fastq.gz
- Allowed file extensions: '.fa', '.fasta', '.fas', '.fna', '.seq', '.zip','.txt','.fastq','.fq','.gz'

Fasta example:

>AF184762

tcctcctcctccactgcacaaggtctctctccccggtcatgctgacgcaatcaccctcta
tttctgcctccctgggagcctcggtcaacctcacctgcactctgaccagtgggcacagac
gttacgccatcgcatggcatcagcaattgtcagggaagggccctcgtttcttgatgagac
ttaacagtgatggcacttacaccaggggggacgggattcctgatcgcttctccggctcca
cctctgggcctgagcgctacctcaccatctccagcctccagtctgaagatgaggcagatt
attactgtcagacctggggcactggcctttgggttttcggcggagggaccagtctgaccg
tcttaggtcagcccaaggctgcccctcg

Fastq Example

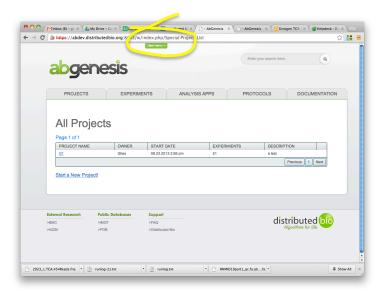
@EAS54_6_R1_2_1_413_324 CCCTTCTTGTCTTCAGCGTTTCTCC + ;;3;;;;;7;;;;88 @EAS54_6_R1_2_1_540_792 TTGGCAGGCCAAGGCCGATGGATCA + ;;;;;;;7;;;;-;;;3;83 @EAS54_6_R1_2_1_443_348 GTTGCTTCTGGCGTGGGTGGGGGGG +EAS54_6_R1_2_1_443_348 ;;;;;;;;9;7;;7;393333

Not Supported: Microsoft Word; Excel.

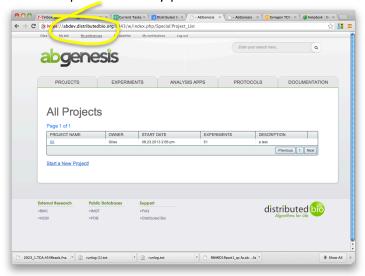


Appendix B - Change Your Password

From any screen pull down the User menu from the top of the screen:

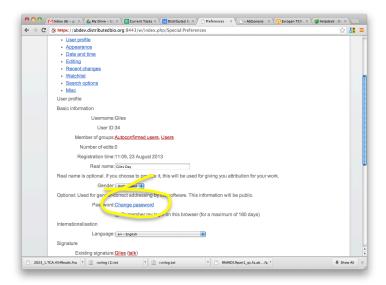


From the options click My preferences

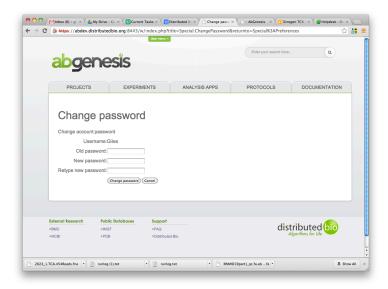




The Preferences page will load. Scroll down until you Change password, click on that link.



Fill out the form and click the change password button



The next time you log into the system you will need to use this password. If you forget your password contact your system administrator to reset it.



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Appendix C - File Types

file1_qc.fa.abgen-

Your input file after it has been through the Quality Control file1_qc.fa process file1_qc.fa.abgen.aa.VDJ.H3.L3. Protein fasta file with extended headers, seq_id and CH1.dnaH3.a2m transslation frame in position one of header. Aligned protein sequence file1 gc.fa.abgen.aa.VDJ.H3.L3. Protein fasta file with extended headers. Same headers as CH1.dnaH3.fa .VDJ.H3.L3.CH1.dnaH3.fa. First field in header is seq_id and translation frame Protein fasta file. Extended headers with developability file1 qc.fa.abgen.developabilityannotated.fa annotations. file1_qc.fa.abgen.fa Original input file with unique identifiers applied file1_qc.fa.abgen.unclass.fa All sequences that did not match a V,D or J gene file1 qc.fa.abgen.VDJ.H3.L3.CH Output of VDJFasta. DNA fasta file with extended header 1.dnaH3.fa with format: >seq_id;vh_field;dh_field;jh_field;h3;l3;isotype;np_boundari es;h1;h2;l1;l2;vl_field;jl_field;dna_h3 file1_qc.fa.abgen.VDJ.H3.L3.CH Sequences and clone tab separated for **Heavy** chain only. 1.dnaH3.fa.clones.txt All sequences in file and will show NO_HIT if they do not match a Heavy chain clone file1 qc.fa.abgen-Tab separated table of every sequence that matched that V[H|K|L].canonical.tab.txt chain and the CDR canonical classifications file1 qc.fa.abgen-Tab separated table of every sequence that matched that V[H|K|L].cdrs.tab.txt chain and the CDRs file1 qc.fa.abgen-V[H|K|L]-DNA fasta file with extended headers, same as clones.fa .VDJ.H3.L3.CH1.dnaH3.fa. Contains all sequences that match the chain in the filename (VH, VK or VL). Summary statistics for each unique clone matching the file1_qc.fa.abgen-V[H|K|L]-clonechain in the file name (VH, VK, VL). Tab separated: Clone table.txt frequency, number of matching seqs, Vgene, Dgene, Jgene, Germline %, CDR1, CDR2, CDR3, Matching seq_ids (, separated) file1_qc.fa.abgen-V[H|K|L]-clone-Full tab separated listing of every sequence and every table.xls property calculated by the analysis pipeline. Can be VERY LARGE file1_qc.fa.abgen-Tab separated table of every sequence that matched that V[H|K|L].frameworks.tab.txt chain and the CDRs

Tab separated table of every sequence that matched that



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V[H|K|L].liabilities.tab.txt chain and their liabilities - there is no summary table in this

file

file1_qc.fa.abgen-Tab separated table of every sequence that matched that V[H|K|L].numbering.tab.txt chain and the Kabat Numbering for that sequence

file1_qc.fa.abgen-Tab separated table of every sequence that matched that

V[H|K|L].shm.tab.txt chain and their Somatic Hyper-mutations